+80/CCR6 5'- ATTTCAGCGATGTTTTCGACTC -3' (SEQ ID NO: 1) forward primer,
-1081/CCR6 5'- GGAGAAGCCTGAGGACTTGTA -3' (SEQ ID NO: 2) reverse primer,
+154/CCR7 5'-GATTACATCGGAGACAACACC -3' (SEQ ID NO: 3)
forward primer and
-1202/CCR7 5'-TAGTCCAGGCAGAAGAGTCG -3' (SEQ ID NO: 4) reverse primer were used for RT-PCR and sequencing. For both chemokine receptors, the reaction mixture was subjected to 30 and 35 cycles of PCR with the following conditions : 94°C for 1 min, 61.5°C for 2 min and 72°C for 3 min. PCR products were visualized on 1.2% agarose gels containing 0.5 µg/ml ethidium bromide. Reaction products migrating at the predicted size (1,021 bp for CCR6 and 1,067 bp for CCR7) were gel purified and subcloned into pCRII TA cloning vector (Invitrogen, Leek, The Netherlands) for sequencing verification on an ABI 373A Sequencer (Applied Biosystems, Foster City, CA.) using dye terminator technology. Two other oligonucleotides,
-622/CCR6 5'-GCTGCCTTGGGTGTTGTATTT -3' (SEQ ID NO: 5)
and
+662/CCR7 5'-AGAGGAGCAGCAGTGAGCAA -3' (SEQ ID NO: 6), were used as probes for hybridization with the PCR products separated on 1.2% agarose gel and blotted onto Hybond N^+ membranes (Amersham, Les Ulis, France).

Please replace the first full paragraph on page 19 with the following rewritten paragraph:

In situ hybridization. *In situ* hybridization was performed as described (Peuchmaur, *et al.*, 1990, *Am. J. Pathol.* **136**:383-390). Two couple primers were used for amplifying by RT-PCR the majority of the open reading frame of MIP-3α (Accession No. D86955)and MIP-3βα (Accession No. U77180) genes.

+77/MIP-3α 5'- TTGCTCCTGGCTGCTTTG -3' (SEQ ID NO: 7) forward primer and

-425/MIP-3α 5'- ACCCTCCATGATGTGCAAG -3' (SEQ ID NO: 8) reverse primer, +25/MIP-3β 5'- CTGCTGGTTCTCTGGACTTC -3' (SEQ ID NO: 9) forward primer and

-439/MIP-3β 5'- CACACTCACACTCACACACAC -3' (SEQ ID NO: 10) reverse primer, were used as described above with an annealing temperature at 62°C. Then, PCR products were cloned into pCRII TA cloning vector (Invitrogen, Leek, The Netherlands) for the generation of sense and anti-sense probes with the adapted promoters. Sense and antisense ^{35}S-labeled probes of MIP-3α and MIP-3β, were obtained by run off transcription of the 367 bp and 435 bp fragments, respectively. Six µm human tonsil sections were fixed in acetone and 4% paraformaldehyde followed by 0.1 M